



Special Issue: Interaction between gut microbiota and host immune cells

Mini Review

Bacteriotherapy reintroduces “old friends” in IBD

Kosaku Nanki¹⁾, Makoto Naganuma²⁾, Shinta Mizuno³⁾,
Katsuyoshi Matsuoka⁴⁾ and Takanori Kanai^{1,*})

¹⁾Division of Gastroenterology and Hepatology, Department of Internal Medicine, Keio University School of Medicine, Shinjuku-ku, Tokyo, Japan

²⁾Center for Diagnostic and Therapeutic Endoscopy, Keio University School of Medicine, Shinjuku-ku, Tokyo, Japan

³⁾Department of Gastroenterology and Hepatology, Saiseikai Central Hospital, Minato-ku, Tokyo, Japan

⁴⁾Department of Gastroenterology and Hepatology, Tokyo Medical and Dental University, Bunkyo-ku, Tokyo, Japan

Various strains of microorganisms inhabit the human gut and greatly impact human health. Recent advances in next-generation sequencing techniques revealed a correlation between alterations to the composition of gastrointestinal microbiota: called dysbiosis, and inflammatory bowel disease (IBD), a chronic inflammatory intestinal disorder comprising ulcerative colitis (UC) and Crohn's disease (CD). These alterations are suspected to be the causes of IBD. Significant lifestyle and environmental changes in modern developed countries may be responsible for the altered gastrointestinal microbiota, and may have greatly contributed to the rapid rise of IBD in the modern era. To date, many trials attempted to treat IBD by restoring altered microbiota using such methods as probiotics and fecal microbiota transplantation (FMT). Currently, more sophisticated and convenient methods of FMT are being devised. The focus is now on FMT, which is expected to be the new direction of IBD therapy.

Rec.2/10/2015, Acc.3/22/2015, pp122-128

*Correspondence should be addressed to:

Takanori Kanai, M.D., Ph.D., Division of Gastroenterology and Hepatology, Department of Internal Medicine, Keio University School of Medicine, 35 Shinanomachi, Shinjuku-ku, Tokyo 160-8582, Japan. Phone: +81-3-5843-7090, Fax: +81-3-5843-7091, E-mail: takagast@z2.keio.jp

Key words fecal microbiota transplantation, probiotics, microbiota, dysbiosis, inflammatory bowel disease

Introduction

Inflammatory bowel disease (IBD), which comprises ulcerative colitis (UC) and Crohn's disease (CD), is characterized by chronic inflammation of the gastrointestinal

tract. IBD patients experience remission and relapse cycles of inflammation, and present with diarrhea, bloody stools, and weight loss. While the cause of IBD remains unknown, more than 160 IBD-associated susceptibility genes have



been identified in multiple studies¹), suggesting that genetic factors probably contribute to IBD. As the most widely used IBD therapy is immunosuppressive therapy, inappropriate immune response is strongly implicated in the pathogenesis of IBD. Environmental factors are considered to mediate genetic factors and inappropriate immune responses. To date, the most consistent pathogenetic mechanism proposed for IBD is an inappropriate immune response against gastrointestinal commensal microbiota triggered by environmental factors²).

The number of IBD patients has shown tremendous increases, especially in developed countries in the modern era. The involvement of alterations to gastrointestinal microbiota in IBD patients has been highlighted as a potential cause. Sellon et al. reported that genetically engineered IBD mice do not develop colitis in germ-free conditions³), suggesting that gastrointestinal microbiota play an important role in triggering IBD inflammations. The human gastrointestinal tract contains 10^{14} bacteria composed of over 1,000 species that build gastrointestinal microbiota⁴). The recent advance of next-generation sequencing techniques has made it possible to exhaustively analyze gastrointestinal microbiota. Using this technique, it was discovered that a disruption of the balance of microbiota known as “dysbiosis” likely contributes to IBD exacerbation⁵). Modern people have developed methods of preservation, transportation, and sewage disposal, promoted hygienic environments, and dramatically changed their diets to include high sugar/low dietary fiber. As a result, modern people have lost the balance of their gastrointestinal microbiota, in a manner, their “old friends”. Given this change, it has become apparent that the number of IBD patients has been increasing gradually with the modernization and westernization of our lifestyles. In this review, we will discuss the correlation between IBD and alterations of gastrointestinal microbiota, as well as bacteriotherapies that reconnect us with our “old friends”.

Lifestyle changes and microbiota

When no freezing or refrigeration technology was available, fermentation was the essential method of food preservation. People of this era frequently ate fermented food, including probiotics (e.g., *Lactobacillus*), and naturally inoculated many types of beneficial bacteria into their gastrointestinal tracts. Along with modernization and industrialization, the development of freezing and refrigeration technology has allowed for long-term food

preservation independent of fermentation. In addition, the development of transportation technology has enabled the year-round availability of fresh meat, fish, and vegetables while the development of sewage disposal systems made our environment more sanitary, allowing us to preserve food without the risk of rotteness. Thus, people have come to prefer fresh food to fermented food, leading to a decline in the consumption of fermented food. Furthermore, the improvement of hygiene in our environments decreases our exposure to numerous bacteria⁶). The components of our diet have also changed dramatically in the modern era. Modern individuals prefer foods that are low in dietary fiber to those that are rich in fiber because of the ease of digestion. Significant reductions in the intake of dietary fibers, which feed microbiota in the human gut, greatly affect the microbiotic environment and reduce the diversity of gastrointestinal microbiota. Additionally, the changes in amount of consumption of fat and other nutrients also lead to dysbiotic state of gut microbiota⁷).

The mode of child delivery and infant feeding method also affect gut microbiota^{8, 9}). Pre-birth, children are in germ-free conditions. They are then exposed to many types of microorganisms at delivery for the first time, and are colonized by commensal microorganisms within 2-3 days after birth. Several beneficial bacteria such as *Lactobacillus* species are present in the birth canal. These beneficial bacteria colonize by ingestion when the baby is delivered vaginally. However, a baby delivered by Caesarean section is not exposed to these bacteria. While human breast milk was traditionally thought to be sterile, it in fact contains many bacteria and is one of the main sources by which bacteria are ingested¹⁰). *Bifidobacterium*, a known probiotic, constitutes a substantial portion of the microbiota in the feces of breast-fed infants¹¹). Human milk may form healthier gut microbiota in infants¹²).

This is the manner in which gastrointestinal microbiota of modern people, especially those in westernized and industrialized countries, has been dramatically altered along with the loss of beneficial bacteria.

Dysbiosis may induce intestinal inflammation in IBD

To date, no specific bacterium that induces IBD has been identified. In the pathophysiology of IBD, the condition is thought to result from the alteration of microbiota (i.e., dysbiosis) rather than from a single specific pathogen¹³). Indeed, several studies reported reduced diversity and total



amounts of gastrointestinal microbiota in IBD patients^{5, 14-16}. The alterations of gastrointestinal microbiota in IBD include reduced numbers of *Firmicutes* and *Bacteroidetes* and increased numbers of *Proteobacteria* and *Actinobacteria*. These changes are more obvious in inflamed sites than in non-inflamed sites, suggesting the possible involvement of dysbiosis in intestinal inflammation. However, it remains uncertain whether these alterations of microbiota in IBD patients are a cause or a consequence of the inflammation. Interestingly, wild-type mice co-housed with genetically engineered mice that developed spontaneous UC-like colitis also developed a similar colitis¹⁷. This suggests that a state of dysbiosis in the gut can affect normal gastrointestinal microbiota and induce a dysbiotic state. Altered microbiota can also promote intestinal inflammation.

Recently, several studies showed correlations between the intestinal immune systems of IBD patients and specific bacterial strains. For example, segmented filamentous bacteria (SFB) are sufficient to induce Th17 cells in the intestine¹⁸. Th17 cells secrete IL-17 and IL-22 and play a crucial role in IBD inflammation. Clostridia ferment dietary fibers and produce short-chain fatty acids (SCFAs) such as acetate, butyrate, and propionate. Butyrate is one of the most significant sources of nutrients for colonocytes, and contributes not only to the proliferation of intestinal epithelium but also to the suppression of intestinal inflammation. Several studies have revealed that butyrate induces the differentiation of regulatory T cells that produce IL-10 and suppress excessive immune responses¹⁹⁻²¹. Furthermore, Atarashi et al. reported that a mixture of 46 mouse-derived *Clostridia* strains promoted transforming growth factor (TGF)- β and induced IL-10-producing regulatory T cells in inoculated germ-free mice²². These results suggest that the decrease of Clostridia observed in IBD patients causes aberrant functioning of regulatory T cells and reduces their numbers, consequently contributing to the development and maintenance of intestinal inflammation. Recently, our group demonstrated that *clostridium butyricum* (CB) prevents acute experimental colitis in mice through induction of IL-10, an anti-inflammatory cytokine²³. CB directly triggered IL-10 production by intestinal macrophages in inflamed mucosa via the TLR2/MyD88 pathway. The colitis-preventing effect of CB was negated in macrophage-specific IL-10-deficient mice, suggesting that induction of IL-10 by intestinal macrophages is crucial for the probiotic action of CB. These results suggested that CB may be useful for IBD patients to

prevent colitis as probiotics.

Restored dysbiosis as a treatment for IBD

As discussed above, the abnormal composition of gastrointestinal microbiota is likely to be deeply involved in the pathophysiology of IBD, and communicated dysbiotic gastrointestinal microbiota can cause the development of intestinal inflammation. Against the background of these findings, several trials have attempted restoring the abnormal microbial composition for the treatment of IBD (Fig. 1).

1) Probiotics

Probiotics are live bacteria that benefit human health. Several clinical trials have demonstrated the efficacy of probiotics for IBD. For example, *Escherichia coli* Nissle 1917 was found to be safe and effective in maintaining remission in UC equivalent to that of mesalazine²⁴. VSL#3, a mixture of 8 strains of bacteria, showed efficacy in the prevention of recurrent pouchitis²⁵. VSL#3 has also been shown to be effective in inducing remission in active UC patients²⁶. There is little evidence for the efficacy of probiotics in CD patients. One systematic review of randomized control trials of probiotics in IBD patients concluded that there is insufficient evidence to recommend probiotic use in CD patients²⁷.

However, two Cochrane reviews evaluated the efficacy of probiotics for the induction or maintenance of remission in UC patients^{28, 29}, finding limited benefits of probiotics in

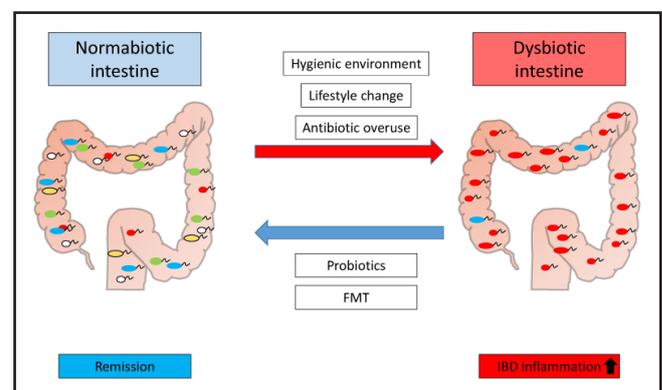


Fig. 1 Treatment restoring altered microbiota in IBD

The overuse of antibiotics, lifestyle changes, and the development of a hygienic environment resulted in the dysbiosis of normal gut microbiota. Dysbiosis may cause intestinal inflammation, and it is thus expected that treating dysbiosis with probiotics or fecal microbiota transplantation will dramatically improve these inflammations.



this population. Therefore, probiotics are not recommended as first-line therapy in IBD. There are three considerable reasons for the limited efficacy of probiotics in IBD.

First, probiotics contain a lower bacterial load compared to the gastrointestinal microbiota. There are only 10^6 - 10^9 bacteria in probiotics, 100,000-fold less bacteria than the amount found in human beings. Thus, the impact of probiotics on gastrointestinal microbiota may be very small.

Second, there is a lack of bacterial diversity in probiotics. Probiotics are composed of only a single strain or a few strains of bacteria. By contrast, the gastrointestinal microbiota is composed of over 1,000 species of bacteria that build a complex web of interactions. A few types of bacterial strains may exert only a small effect against these complex interactions.

Finally, the mechanism of "colonization resistance", whereby ingested pathogens and foreign substances are excluded to maintain health and homeostasis may inhibit probiotic colonization. Healthy hosts with normal immune function are consistently exposed to pathogenic microorganisms, but they rarely develop diseases because of colonization failure due to colonization resistance. Commensal bacteria that build gastrointestinal microbiota have evolved and adapted to fulfill the nutritional niche, and compete with colonizing pathogens³⁰. Indirect mechanisms of colonization resistance that are mediated by the immune system have also been suggested. Commensal bacteria have acquired tolerance to both the immune response and to antimicrobial peptide (AMP)³¹. When pathogenic bacteria invade a host intestine, commensal bacteria induce the host's immune response as well as the production of AMP. Consequently, pathogenic bacteria are eliminated from the intestine before they can adapt to the environment.

Thus, probiotics may have a limited effect on aberrant gastrointestinal microbiota. Probiotics can only ameliorate the symptoms of IBD, but are considered insufficient to strongly induce remission in IBD patients. For maintenance treatment, probiotics may be considered for patients who are intolerant to 5-ASA preparations; however, the underlying evidence is weak to recommend its use in all UC patients, and the specific probiotic is not available in many countries³².

2) Fecal microbiota transplantation

Fecal microbiota transplantation (FMT) is a underexplored treatment geared at restoring aberrant microbial compositions by delivering fecal microbiota derived from healthy

donors into the recipient's gastrointestinal tract. *Clostridium difficile* infection (CDI) is an intestinal disorder that is often associated with a course of antibiotics that causes dysbiosis in the gut. It is difficult to cure patients with recurrent CDI by repeatedly administering vancomycin or metronidazole. A randomized controlled trial comparing FMT with standard vancomycin therapy for recurrent CDI was conducted in the Netherlands³³. Surprisingly, FMT had a much higher cure rate (81%) than standard antibiotic treatment (31%) in these recurrent CDI patients. The fecal microbiota used in FMT are obtained from healthy donors and contain numerous strains of bacteria that are thought to easily adapt to the intestinal environment because they previously colonized the donor intestine.

FMT has been highlighted as a treatment for restoring dysbiosis in IBD. In what was the first implementation of FMT in UC, in 1989, Bennet et al. reported one UC patient with a continuously active intestinal inflammation who received FMT and experienced remission³⁴. Borody et al. reported using consecutive FMT to induce remission in 6 active UC patients, all of whom achieved clinical remission without severe adverse events³⁵.

However, there are several reports of UC patients who received FMT and did not achieve clinical remission³⁶⁻³⁸. Kump et al. reported that patients whose microbiota dramatically changed to resemble the composition of the donor fecal microbiota did not achieve clinical remission. By contrast, Angelberger et al. reported that the successful inoculation of fecal microbiota from donor feces into the recipient intestine correlated with the successful achievement of clinical remission.

A systematic review published in 2014 included 18 studies of FMT for IBD and a total of 122 patients. According to the results, 54 of 119 patients (45%) achieved clinical remission without severe adverse events³⁹. However, publication bias should be considered because of the inclusion of 8 case studies. Supported by these backgrounds, we conducted a clinical trial treating IBD with using FMT in Japan⁴⁰.

There are several obstacles to FMT treatment. FMT transfers not only beneficial bacteria, but also pathogenic microorganisms such as hepatitis B virus, hepatitis C virus, human immunodeficiency virus, *Helicobacter pylori*, and pathogenic amoebae. Therefore, donors must be screened carefully. Despite careful screening, the risk of transmitting unknown pathogens remains⁴¹. FMT may also transfer extra-intestinal disorders that have significant correlations with alterations of intestinal microbiota such



as type 2 diabetes mellitus, multiple sclerosis, chronic fatigue syndrome, and idiopathic thrombocytopenic purpura⁴²). In addition, such donor screenings are time consuming, making FMT unsuitable for the patient with severe disease activity requiring immediate intervention.

To resolve these issues, the feces bank named “Open Biome” was established in the US in 2012. A recent report showed that frozen FMT inocula have equivalent efficacy for treating CDI patients⁴³). The “Open Biome” pools frozen feces obtained from donors proven safe via screening. Owing to the “Open Biome”, the application of FMT to the treatment of dysbiosis is easier, quicker, and safer.

Future perspectives

The routes of infusion of fecal material vary across trials and include administration by nasogastric tube, nasojejunal tube, esophagogastroduodenoscopy, colonoscopy, or retention enema. Because FMT is a developing treatment, the optimal administration route is uncertain. FMT protocols are still complicated, and a more convenient method is needed. Youngster et al. conducted an open-label, single-group trial of a less invasive and easier to use FMT treatment in which they orally administered frozen encapsulated fecal material from unrelated donors to relapsing CDI patients. They reported no serious adverse events and a 90% rate of clinical resolution⁴⁴). In Europe and the United States, an ongoing trial is investigating the use of “Artificial Feces” that contain large amounts of various probiotics instead of FMT⁴⁵). “Artificial Feces” excel in terms of cost, stability of effectiveness, and ethicality. These advantages facilitate the treatment's commercialization for restoring dysbiosis.

Recent reports and our study found that specific strains of bacteria suppress intestinal inflammation in a gnotobiotic murine model^{23, 46}). In this manner, microorganisms that can potentially resolve inflammation are being identified. The future progress of research on bacteria and bacterial interactions that correlate with intestinal inflammation may identify optimal combinations of probiotics that will enable the formulation of ideal “Artificial Feces” that can restore dysbiosis much more easily, safely, and effectively.

Conclusion

As discussed above, the numbers of IBD patients in developed countries are rapidly increasing along with alterations of the composition of gastrointestinal microbiota. FMT may have great potential for restoring dysbiosis and

treating IBD. However, there is limited evidence regarding FMT for IBD, and better optimized and more sophisticated methods of FMT are needed.

Feces are generally viewed as “dirty” materials, however, the feces of healthy individuals are actually a “clean drug” that contains an enormous number of beneficial bacteria and can restore the disrupted balance of microbiota. It is important to reacquaint the gastrointestinal tract with these “old friends”, and to maintain a “clean” gastrointestinal tract in the management of IBD. Therefore, further insights into probiotics should be obtained through cooperation not only between clinicians and drug companies, but also with the food industry.

Acknowledgment and Source of funding

The authors thank the members of the Keio University IBD team: Haruhiko Ogata, Yasushi Iwao, Nagamu Inoue, Tadakazu Hisamatsu, Tomoharu Yajima, Yoshihiro Nakazato, Tatsuya Takeshita, Keiichiro Saigusa, Kozue Takeshita, Kiyoto Mori, Mari Arai, Shinya Sugimoto, and Hirota Kiyohara. This study was supported by Grants-in-Aid from the Japanese Ministry of Education, Culture, Sports, Science and Technology; the Health and Labour Sciences Research Grants for Research on Intractable Diseases from the Ministry of Health, Labour and Welfare of Japan; and the Keio University Medical Fund.

Conflict of interests

K.M. received a research grant, lecture fees, and consulting fees from Mitsubishi Tanabe Pharma Corporation, Tsumura & Co., Ltd., Asahi Kasei Medical Co., Ltd., Eisai Co., Ltd., Abbvie, JIMRO Co., Ltd., Kyorin Pharmaceutical Co., Ltd., and ZERIA Co., Ltd. T.K. received a research grant, lecture fees, and consulting fees from Mitsubishi Tanabe Pharma Corporation, Takeda Pharmaceutical Co., Ltd., Ltd., Asahi Kasei Medical Co., Ltd., JIMRO Co., Ltd., Kyorin Pharmaceutical Co., Ltd., and ZERIA Co., Astellas Pharma Inc., Pfizer Inc., Biofermin pharmaceutical Co., Ltd., Daiichi Sankyo Co., Ltd., Ajinomoto Co., Inc., Otsuka pharmaceutical Co., Ltd., Eidia Co., Ltd., and Public Health Research Foundation.

Reference

- 1) Jostins L, Ripke S, Weersma RK, et al: Host-microbe interactions have shaped the genetic architecture of inflammatory bowel disease. *Nature*. 2012; 491: 119-24.
- 2) Sartor RB: Mechanisms of disease: pathogenesis of Crohn's disease and ulcerative colitis. *Nat Clin Pract Gastroenterol Hepatol*. 2006; 3: 390-407.
- 3) Sellon RK, Tonkonogy S, Schultz M, et al: Resident enteric bacteria are necessary for development of



- spontaneous colitis and immune system activation in interleukin-10-deficient mice. *Infect Immun.* 1998; 66: 5224-5231.
- 4) Qin J, Li R, Raes J, Arumugam M, et al: A human gut microbial gene catalogue established by metagenomic sequencing. *Nature.* 2010; 464: 59-65.
 - 5) Manichanh C, Rigottier-Gois L, Bonnaud E, et al: Reduced diversity of faecal microbiota in Crohn's disease revealed by a metagenomic approach. *Gut.* 2006; 55: 205-211.
 - 6) Guarner F: Hygiene, microbial diversity and immune regulation. *Curr Opin Gastroenterol.* 2007; 23: 667-672.
 - 7) Goldsmith JR, Sartor RB: The role of diet on intestinal microbiota metabolism: downstream impact on host immune function and health, and therapeutic implications. *J Gastroenterol.* 2014; 49: 785-798.
 - 8) Penders J, Thijs C, Vink C, et al: Factors influencing the composition of the intestinal microbiota in early infancy. *Pediatrics.* 2006; 118: 511-521.
 - 9) Biasucci G, Benenati B, Morelli L, et al: Cesarean delivery may affect the early biodiversity of intestinal bacteria. *J Nutr.* 2008; 138: 1796S-1800S.
 - 10) Heikkilä MP, Saris PE: Inhibition of *Staphylococcus aureus* by the commensal bacteria of human milk. *J Appl Microbiol.* 2003; 95: 471-478.
 - 11) Bezirtzoglou E, Tsiotsias A, Welling GW: Microbiota profile in feces of breast- and formula-fed newborns by using fluorescence in situ hybridization (FISH). *Anaerobe.* 2011; 17: 478-482.
 - 12) Asakuma S, Hatakeyama E, Urashima T: Physiology of consumption of human milk oligosaccharides by infant gut-associated bifidobacteria. *J Biol Chem.* 2011; 286: 34583-34592.
 - 13) Sartor RB: Microbial influences in inflammatory bowel diseases. *Gastroenterology.* 2008; 134: 577-594.
 - 14) Frank DN, St Amand AL, Feldman RA, et al: Molecular-phylogenetic characterization of microbial community imbalances in human inflammatory bowel diseases. *Proc Natl Acad Sci U S A.* 2007; 104: 13780-13785.
 - 15) Ott SJ, Musfeldt M, Wenderoth DF, et al: Reduction in diversity of the colonic mucosa associated bacterial microflora in patients with active inflammatory bowel disease. *Gut.* 2004; 53: 685-693.
 - 16) Takaishi H, Matsuki T, Nakazawa A, et al: Imbalance in intestinal microflora constitution could be involved in the pathogenesis of inflammatory bowel disease. *Int J Med Microbiol.* 2008; 298: 463-472.
 - 17) Garrett WS, Lord GM, Punit S, et al: Communicable ulcerative colitis induced by T-bet deficiency in the innate immune system. *Cell.* 2007; 131: 33-45.
 - 18) Ivanov II, Atarashi K, Manel N, et al: Induction of intestinal Th17 cells by segmented filamentous bacteria. *Cell.* 2009; 139: 485-498.
 - 19) Furusawa Y, Obata Y, Fukuda S, et al: Commensal microbe-derived butyrate induces the differentiation of colonic regulatory T cells. *Nature.* 2013; 504: 446-450.
 - 20) Arpaia N, Campbell C, Fan X, et al: Metabolites produced by commensal bacteria promote peripheral regulatory T-cell generation. *Nature.* 2013; 504: 451-455.
 - 21) Smith PM, Howitt MR, Panikov N, et al: The microbial metabolites, short-chain fatty acids, regulate colonic Treg cell homeostasis. *Science.* 2013; 341: 569-573.
 - 22) Atarashi K, Tanoue T, Shima T, et al: Induction of colonic regulatory T cells by indigenous *Clostridium* species. *Science.* 2011; 331: 337-341.
 - 23) Hayashi A, Sato T, Kamada N, et al: A single strain of *Clostridium butyricum* induces intestinal IL-10-producing macrophages to suppress acute experimental colitis in mice. *Cell Host Microbe.* 2013; 13: 711-722.
 - 24) Kruis W, Frick P, Pokrotnieks J, et al: Maintaining remission of ulcerative colitis with the probiotic *Escherichia coli* Nissle 1917 is as effective as with standard mesalazine. *Gut.* 2004; 53: 1617-1623.
 - 25) Mimura T, Rizzello F, Helwig U, et al: Once daily high dose probiotic therapy (VSL#3) for maintaining remission in recurrent or refractory pouchitis. *Gut.* 2004; 53: 108-114.
 - 26) Tursi A, Brandimarte G, Papa A, et al: Treatment of relapsing mild-to-moderate ulcerative colitis with the probiotic VSL#3 as adjunctive to a standard pharmaceutical treatment: a double-blind, randomized, placebo-controlled study. *Am J Gastroenterol.* 2010; 105: 2218-2227.
 - 27) Ghouri YA, Richards DM, Rahimi EF, et al: Systematic review of randomized controlled trials of probiotics, prebiotics, and synbiotics in inflammatory bowel disease. *Clin Exp Gastroenterol.* 2014; 7: 473-487.
 - 28) Naidoo K, Gordon M, Fagbemi AO, et al: Probiotics for maintenance of remission in ulcerative colitis. *Cochrane Database Syst Rev.* 2011; (12): CD007443.
 - 29) Mallon P, McKay D, Kirk S, et al: Probiotics for induction of remission in ulcerative colitis. *Cochrane Database Syst Rev.* 2007; (4): CD005573.



- 30) Lawley TD, Walker AW: Intestinal colonization resistance. *Immunology*. 2013; 138: 1-11.
- 31) Cullen TW, Schofield WB, Barry NA, et al: Gut microbiota. Antimicrobial peptide resistance mediates resilience of prominent gut commensals during inflammation. *Science*. 2015; 347: 170-175.
- 32) Naganuma M, Sakuraba A, Hibi T: Ulcerative colitis: prevention of relapse. *Expert Rev Gastroenterol Hepatol*. 2013; 7: 341-351.
- 33) van Nood E, Vrieze A, Nieuwdorp M, et al: Duodenal infusion of donor feces for recurrent *Clostridium difficile*. *N Engl J Med*. 2013; 368: 407-415.
- 34) Bennet JD, Brinkman M: Treatment of ulcerative colitis by implantation of normal colonic flora. *Lancet*. 1989; 1: 164.
- 35) Borody TJ, Warren EF, Leis S, et al: Treatment of ulcerative colitis using fecal bacteriotherapy. *J Clin Gastroenterol*. 2003; 37: 42-47.
- 36) Kump PK, Gröchenig HP, Lackner S, et al: Alteration of intestinal dysbiosis by fecal microbiota transplantation does not induce remission in patients with chronic active ulcerative colitis. *Inflamm Bowel Dis*. 2013; 19: 2155-2165.
- 37) Suskind DL, Singh N, Nielson H, et al: Fecal microbial transplant via nasogastric tube for active pediatric ulcerative colitis. *J Pediatr Gastroenterol Nutr*. 2015; 60: 27-29.
- 38) Angelberger S, Reinisch W, Makristathis A, et al: Temporal bacterial community dynamics vary among ulcerative colitis patients after fecal microbiota transplantation. *Am J Gastroenterol*. 2013; 108: 1620-1630.
- 39) Colman RJ, Rubin DT: Fecal microbiota transplantation as therapy for inflammatory bowel disease: a systematic review and meta-analysis. *J Crohns Colitis*. 2014; 8: 1569-1581.
- 40) Matsuoka K, Mizuno S, Hayashi A, et al: Fecal microbiota transplantation for gastrointestinal diseases. *Keio J Med*. 2014; 63: 69-74.
- 41) El-Matary W, Simpson R, Ricketts-Burns N: Fecal microbiota transplantation: are we opening a can of worms? *Gastroenterology*. 2012; 143: e19.
- 42) Smits LP, Bouter KE, de Vos WM, et al: Therapeutic potential of fecal microbiota transplantation. *Gastroenterology*. 2013; 145: 946-53.
- 43) Youngster I, Sauk J, Pindar C, et al: Fecal microbiota transplant for relapsing *Clostridium difficile* infection using a frozen inoculum from unrelated donors: a randomized, open-label, controlled pilot study. *Clin Infect Dis*. 2014; 58: 1515-1522.
- 44) Youngster I, Russell GH, Pindar C, et al: Oral, capsulized, frozen fecal microbiota transplantation for relapsing *Clostridium difficile* infection. *JAMA*. 2014; 312: 1772-1778.
- 45) Petrof EO, Gloor GB, Vanner SJ, et al: Stool substitute transplant therapy for the eradication of *Clostridium difficile* infection: 'RePOOPulating' the gut. *Microbiome*. 2013; 1: 3.
- 46) Nishikawa H, Fukuda S, Saito T, et al: Treg induction by a rationally selected mixture of *Clostridia* strains from the human microbiota. *Nature*. 2013; 500: 232-236.